FLAVONOID AGLYCONES OF HOLOCARPHA OBCONICA*

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Abstract—Eleven flavonoid aglycones were isolated from the dichloromethane leaf-wash of the tarweed, *Holocarpha obconica*. Included within the extract were three novel flavanones, 7,4'-dimethoxyflavanone, 5,6,3',4'-tetrahydroxy-7-methoxy-flavanone, and 3',4'-dihydroxy-3,7-dimethoxyflavanone.

INTRODUCTION

The genus Holocarpha (DC.) Greene (Heliantheae: Madiinae) consists of four species of summer-flowering, self-incompatible annuals restricted to central and southern California [1]. Its species possess low chromosome numbers (N = 4 and 6) and highly specialized leaf and trichome anatomy [1-3], suggesting an advanced evolutionary disposition for the genus within the subtribe. These tarweeds are copiously glandular, and exude considerable amounts of aromatic resin. No previous reports of flavonoids from Holocarpha have appeared. Related tarweeds in the genus Hemizonia contain mixtures of flavanones, flavones, and flavonols in their leaf-wash extracts [4]. Emerson et al. [5] found a series of 8methoxylated flavonols in the tarweed, Calycadenia ciliosa Greene, a structural type also found in Hemizonia [4]. However, flavanones were not among the constituents of C. ciliosa.

Holocarpha obconica (Clausen and Keck) Keck contains a complex mixture of flavonoids, including flavanones, flavones, and flavonois. Three of the flavanones, 7,4'-dimethoxyflavanone, 5,6,3',4'-tetrahydroxy-7-methoxyflavanone, and 3'4'-dihydroxy-3,7-dimethoxyflavanone, appear to be new natural products.

RESULTS AND DISCUSSION

Eleven flavonoids were isolated from the dichloromethane leaf-wash of *Holocarpha obconica*. Five flavanones (1-5), two flavones (6, 11), and four flavonols (7-10) were present in the dichloromethane leaf-wash.

The flavanones (1-4) appeared as faint dark spots under UV $_{366\,\mathrm{nm}}$. Compound 1 was found in relatively small quantities. It has the highest R_f of all the flavonoids in H. obconica. It became pale green after spraying with Naturstoffreagenz A (NA). Its UV spectrum was typical of a flavanone, with a shoulder at 332 nm and a peak at 286 nm. Owing to persistent contamination from compound 11, reactions of 1 with the shift reagents were inconclusive. The MS showed a parent ion at m/z=284, indicating the presence of two methoxyl and no hydroxyl groups [6]. The lack of hydroxyl groups, particularly at C-5, is unusual, but not unprecedented [7], in the

composites. Crawford and Stuessy [8] point out that resorcinol precursors are used in the biosynthetic pathways of members of the tribe Heliantheae. Thus, the synthesis of 5-unsubstituted compounds by tarweeds could be expected. The 7,4'-dihydroxyl counterpart of 1, liquiritigenin, has been found in various plants [7], and 6-C-methyl-7,4'-dimethoxyflavanone is known from Adina [9].

Compound 2 is similar to 1, with the addition of a hydroxyl group at C-5. It gave a bluish-green colour with NA and its UV and MS properties conform closely to those reported by Lam and Wrang [10].

Compound 3 became bright red after spraying with NA, suggesting an eriodictyol derivative. Sodium acetate fails to produce a shift relative to its methanol UV spectrum, placing a methoxyl group at C-7. Other shift reagents produce reactions similar to those with eriodictyol [11]. The MS data are in accord with the structure of this compound as 7-methyleriodictyol (M⁺ = 302; A-ring fragment = 167; B-ring fragments = 109 and 136).

Compound 4 became yellow-brown after spraying with NA. A large sodium methoxide shift, lack of a sodium acetate shift, and a slightly acid-labile aluminium chloride shift relative to the UV spectrum in methanol suggest that this flavanone has free hydroxyl groups at C-5, C-3', and C-4', and a methoxyl group at C-7. Its mass spectrum (parent ion, m/z = 318) indicated that a fourth hydroxyl group is located on the A-ring, either at C-6 or C-8. Given the structures of the other compounds characterized in H. obconica, and on the basis of biosynthetic considerations, the most likely structure for 4 is 5,6,3',4'-tetrahydroxy-7-methoxyflavanone. Insufficient material was available for NMR analysis.

Compound 5 is yellow under UV light which is characteristic of flavonols and 5-deoxy-flavanones [11], and became orange on spraying with NA. The methanol UV spectrum of this compound (shoulder at 332 nm and peak at 284 nm) suggests it as a 5-deoxyflavanone. It shows a large sodium methoxide shift, no sodium acetate shift, and an acid-labile aluminium chloride shift. This evidence supports the lack of C-5 substitution, the presence of ortho-dihydroxyl substitution on the B-ring, and the lack of a free hydroxyl group at C-7. Its mass spectrum (parent ion, m/z = 316) suggested the presence of two methoxyl and two hydroxyl groups. The fragmentation pattern of this compound agreed with the placement of a methoxyl group at C-3 [cf. 12 and 13]. Compound 5 is therefore

^{*}Chemosystematic studies of the tarweeds (Asteraceae: Heliantheae: Madiinae) No. 2.

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3',4'-dihydroxy-3,7-dimethoxyflavanone.

Compound 6 absorbs UV, and is yellow after spraying with NA. Its UV and MS characteristics indicate that it is luteolin 7-methyl ether. Compound 7 (3-methylkaempferol) absorbs UV and is bright green when sprayed with NA. Its UV and MS characteristics coincide with previously published reports [14, 15]. Compound 8 is yellow under UV, and is yellow-orange after spraying with NA. UV and MS data indicate that it is 3,5,3',4'-tetrahydroxy-7-methoxyflavone (rhamnetin). Compound 9 is dark under UV, and orange after spraying with NA. It is similar to 8 in its UV spectrum and reactions with shift reagents and its UV characteristics conform closely to those described by Urbatsch et al. [16] for 5,3',4'-trihydroxy-3,7-dimethoxyflavone. The MS data are also in accord with this structure. Compound 10 absorbs UV, and is orange after spraying with NA. Its UV and MS characteristics correspond closely to previously published 5, 6, 3', 4'-tetrahydroxy-3,7-[17, 18] of reports dimethoxyflavone.

Compound 11 occurs at very low concentrations, and the proposed structure presented here is based largely on MS data. It was found as a contaminant with 1, having a slightly lower R_f in the ethyl formate solvent system. The mass fragment structure reported for eupatorin [19] is in accord with fragments present in the MS of 11.

EXPERIMENTAL

A voucher of Holocarpha obconica from Deer Valley Rd., California is deposited in UC (Ornduff 8917). Air-dried leaves and stems (360 g dry wt) were soaked in CH₂Cl₂. This extract was placed on a Polyclar AT column, and the constituent compounds eluted initially with 3:1 CH₂Cl₂-MeOH, and subsequently increasing proportions of MeOH in the eluent mixture. The compounds were viewed under UV_{366 nm} before and after spraying with Naturstoffreagenz A. Each flavonoid was purified using TLC on Polyamide 6.6 in ethyl formate-cyclohexane-nbutyl acetate-formic acid (50:25:23:2). UV spectra and MS were obtained for the purified compounds.

7,4'-Dimethoxyflavanone (1). UV $\lambda_{\max}^{\text{MeOH}}$ nm: 286, 332sh. MS m/z: 284 [M] +, 177 [A] +, 164 [A - CH₂], 121 [B + CH₂], 107 [B] +.

5,6,3',4'-Tetrahydroxy-7-methoxyflavanone (4). UV \(\times \) MsOH nm: 288, 336sh; NaOMe: 288, 354; NaOAc: 288, 336sh; AlCl₃: 315, 375sh; AlCl₃-HCl: 312, 375sh. MS m/z: 318 [M]⁺, 289 [M - CHO], 209 [A]⁻, 196 [A - CH], 179 [A - OMe], 167 [A - CH - OMe], 123 [B + CH₂], 110 [B]⁺.

3',4'-Dihydroxy-3,7-dimethoxyflavanone (5). UV \(\hat{\text{MeOH}}\) nm:

284, 332sh; NaOMe: 295, 369; NaOAc: 284, 332sh; AlCl₃: 293, 366; AlCl₃-HCl: 293, 366sh. MS m/z: 316 [M] $^+$, 315 [M - H], 287 [M - CHO], 273 [M - C - OMe], 167 [B + C₂H₂ + OMe], 149 [A] $^+$, 137 [B] $^+$, 123 [A - CO].

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REFERENCES

- Munz, P. and Keck, D. D. (1959) A California Flora. University of California Press, Berkeley.
- 2. Johansen, D. A. (1933) Bot. Gaz. 95, 177.
- 3. Carlquist, S. (1959) Am. J. Botany 46, 300.
- Proksch, P., Budzikiewicz, H., Tanowitz, B. D. and Smith, D. M. (1984) Phytochemistry 23, 679.
- Emerson, J. K., Carr, R. L., McCormick, S. and Bohm, B. A. (1986) Biochem. Syst. Ecol. 14, 29.
- Wollenweber, E. and Dietz, V. H. (1979) Phytochem. Bull. 12, 48
- Wollenweber, E. and Dietz, V. H. (1981) Phytochemistry 20, 869
- Crawford, D. J. and Stuessy, T. F. (1981) Am. J. Botany 68, 107
- Srivastava, S. K. and Gupta, R. K. (1983) Ind. J. Chem. 22B, 1064.
- 10. Lam, J. and Wrang, P. (1975) Phytochemistry 14, 1621.
- Mabry, T. J., Markham, K. R. and Thomas, M. B. (1970) The Systematic Identification of Flavonoids. Springer, New York.
- Balza, F. and Towers, G. H. N. (1984) Phytochemistry 23, 2333.
- do Nascimento, M. C. and Mors, W. B. (1981) Phytochemistry 20, 147.
- Valesi, A. G., Rodriguez, E., Vander Velde, G. and Mabry, T.
 J. (1972) Phytochemistry 11, 2821.
- Bacon, J. D., Urbatsch, L. E., Bragg, L. H., Mabry, T. J., Neuman, P. and Jackson, D. W. (1978) Phytochemistry 17, 1939.
- Urbatsch, L. E., Mabry, T. J., Miyakado, M., Ohno, N. and Yoshioka, H. (1976) Phytochemistry 15, 440.
- Shen, M. C., Rodriguez, E., Kerr, K. and Mabry, T. J. (1976) Phytochemistry 15, 1045.
- Ulubelen, A., Kerr, K. M. and Mabry, T. J. (1980) *Phytochemistry* 19, 1761.
- Timmermann, B. N., Mues, R., Mabry, T. J. and Powell, A. M. (1979) Phytochemistry 18, 1855.